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Perinadine A, a Novel Tetracyclic Alkaloid from Marine-Derived Fungus *Penicillium citrinum*

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ABSTRACT

A novel tetracyclic alkaloid, perinadine A (1), was isolated from the cultured broth of the fungus *Penicillium citrinum*, which was separated from the gastrointestine of a marine fish, and the structure was elucidated on the basis of spectroscopic data including 2D NMR spectra. Biogenetically, perinadine A (1) may be derived from citrinin (4), a well-known mycotoxin, and a scalusamide A-type pyrrolidine alkaloid.

Marine-derived fungi of the genus *Penicillium* have proven to be a rich source of structurally unique and biologically active secondary metabolites.¹ In our search for new metabolites from marine-derived fungi,² three new pyrrolidine alkaloids, scalusamides A (2), B, and C, were isolated from the cultured broth of the fungus *Penicillium citrinum*, which was separated from the gastrointestine of an Okinawan marine parrot fish. Recently, a novel tetracyclic alkaloid, perinadine A (1), was isolated from the same fungus, and the structure was elucidated on the basis of spectroscopic

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data including 2D NMR spectra. In this paper, we describe the isolation and structure elucidation of 1.

The fungus *Penicillium citrinum* (strain N055) was separated from the gastrointestine of a parrot fish *Scalus ovifrons* collected at Hedo Cape, Okinawa Island, and grown in PYG liquid medium containing seawater for 10 days at 25 °C. The supernatant of the culture broth (12 L) was extracted

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Table 1. ¹H and ¹³C NMR Data of Perinadine A (1) in CDCl₃

$\begin{array}{cccccccccccccccccccccccccccccccccccc$	position		$\delta_{ m C}$		$\delta_{ m H}$	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1	70.14	70.06^a	$_{\mathrm{CH}}$	4.24 (d, 4.8)	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	3	77.68	77.58	CH	3.75 (m)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	4	37.18	37.15	CH	2.82 (dq, 7.2, 7.2)	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	4a	144.57		\mathbf{C}		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	5	120.11	120.15	C		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	6	160.98	160.99	\mathbf{C}		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	6-OH				12.241 (s)	$12.235^{a} (s)$
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	7	99.96		\mathbf{C}		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	8	147.26		\mathbf{C}		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	8a	114.26		\mathbf{C}		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	9	21.29	21.31	CH_3	$1.34^b (d, 6.7)$	$1.33^b (d, 6.7)$
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	10	18.80		CH_3	$1.22^b (d, 7.2)$	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	11	11.55		-	$2.17^{b} (s)$	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	12	170.78	170.75	C		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$					11.52 (brs)	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$			86.87			5.63 (d, 5.4)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		45.92			2.62 (m)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	4'	28.92	28.98	CH_2	, ,	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$						
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	5'	45.44	45.56	CH_2	, ,	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$					3.61 (m)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$						
$\begin{array}{cccccccccccccccccccccccccccccccccccc$					3.61 (m)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$						
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	9'	38.82	39.29	CH_2		$2.54^c (\mathrm{m})$
$\begin{array}{cccccccccccccccccccccccccccccccccccc$						
12' 32.27 32.25 CH ₂ 1.95 ^c (m) 13' 130.81 130.75 CH 5.33 (m)				_	, ,	
13' 130.81 130.75 CH 5.33 (m)		28.78		_		
` ,				_		
14' 125 16 125 28 CH 5 38 (m)		130.81	130.75		, ,	
` ,	14'	125.16	125.28	CH	5.38 (m)	_
				-		$1.59^b (d, 4.8)$
16' 13.25 13.44 CH_3 1.44 b (d, 6.7) 1.47 b (d, 6.7)	16′	13.25	13.44	CH_3	$1.44^b (d, 6.7)$	$1.47^b (d, 6.7)$

^a These columns were due to minor signals. ^b 3H. ^c 2H.

with EtOAc, and the EtOAc-soluble portions were subjected to silica gel and amino silica gel column chromatographies followed by C₁₈ HPLC (CH₃CN/H₂O/CF₃CO₂H) to afford perinadine A (1, 5.1 mg). From the other fractions of the EtOAc-soluble portions, scalusamides A³ (2, 10.6 mg), B (1.0 mg), and C (0.9 mg) and a known pyrrolo[2,1-*b*]oxazine compound 3⁴ and its dihydro congener⁵ were isolated.

Perinadine A^6 (1) was obtained as an optically active colorless solid $\{[\alpha]^{22}_D - 33^\circ (c\ 1.0, CHCl_3)\}$. The molecular formula of 1 was revealed to be $C_{28}H_{37}NO_7$ by negative-mode HRESIMS $[m/z\ 498.2510,\ (M-H)^+,\ -0.7\ mmu]$. The IR spectrum suggested the presence of OH/NH (3405 cm⁻¹), carboxylic acid (3300–2700 cm⁻¹), and carbonyl

group(s) (1719 and 1650 cm⁻¹). The ¹³C NMR (Table 1) spectrum of 1 disclosed two sets of 28 carbon signals due to a ketone ($\delta_{\rm C}$ 206.52 and 206.75), two carbonyl ($\delta_{\rm C}$ 170.97 and 170.93; $\delta_{\rm C}$ 170.78 and 170.75), six sp² quaternary carbons ($\delta_{\rm C}$ 160.98 and 160.99; $\delta_{\rm C}$ 147.26; $\delta_{\rm C}$ 144.57; 120.11 and 120.15; both $\delta_{\rm C}$ 114.26; both $\delta_{\rm C}$ 99.96), two sp² methines $(\delta_{\rm C}\ 130.81\ {\rm and}\ 130.75;\ \delta_{\rm C}\ 125.16\ {\rm and}\ 125.28),\ {\rm six}\ {\rm sp}^3$ methines ($\delta_{\rm C}$ 86.75 and 86.87; $\delta_{\rm C}$ 77.68 and 77.58; $\delta_{\rm C}$ 70.14 and 70.06; $\delta_{\rm C}$ 45.44 and 45.56; both $\delta_{\rm C}$ 45.92; $\delta_{\rm C}$ 37.18 and 37.15), the former three of which seemed to be adjacent to a heteroatom, six sp³ methylenes ($\delta_{\rm C}$ 45.44 and 45.56; $\delta_{\rm C}$ 38.82 and 39.29; δ_C 32.27 and 32.25; δ_C 28.92 and 28.98, $\delta_{\rm C}$ 28.78 and 28.92; $\delta_{\rm C}$ 23.00 and 22.94), and five methyls $(\delta_{\rm C} 21.29 \text{ and } 21.31; \text{ both } \delta_{\rm C} 18.80; \delta_{\rm C} 17.91; 12.78 \text{ and}$ 13.44; δ_C 11.55 and 11.50). The chemical shifts for each set of carbon signals of perinadine A (1) were close to each other, probably due to a mixture of epimers at a chiral center (C-7') like scalusamides.3 In the deuterium-induced shift experiment of the ¹³C NMR in CDCl₃ by addition of D₂O, the relatively large shifts were observed for a set of carbons resonated at $\delta_{\rm C}$ 170.78 ($\Delta\delta$ 0.19) and 170.75 ($\Delta\delta$ 0.18) and $\delta_{\rm C}$ 160.98 ($\Delta\delta$ 0.23) and 160.99 ($\Delta\delta$ 0.22), thus suggesting the presence of a carboxylic acid and a phenol group. On the other hand, for the relatively low-field sp² quaternary carbon at δ_C 147.26 was shown a small induced-shift, thus indicating that this carbon was involved in an ether linkage.

The 1H NMR (Table 1) spectrum included two D₂O-exchangeable protons (δ_H 12.241 and 12.235; δ_H 11.52), in which the lowest-field signal was observed as a sharp singlet, indicating the presence of intramolecular hydrogen bonding. Proton and carbon signals for 1 were assigned by detailed analyses of the HMQC spectrum of 1.

The gross structure of perinadine A (1) was elucidated by spectroscopic data including 2D NMR data such as ${}^{1}H^{-1}H$ COSY, NOESY, and HMBC spectra. Four proton networks from H-1 to H-2' and H₂-5', from H-3 to H-4, H₃-9, and H₃-10, from H-7' to H₃-16', and from H₂-9 to H₃-15 were suggested by analysis of the ${}^{1}H^{-1}H$ COSY spectrum (Figure

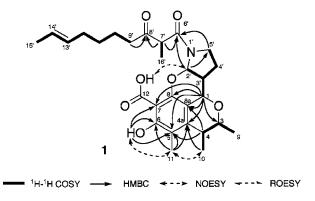


Figure 1. Selected 2D NMR correlations for perinadine A (1).

1). The presence of an *E*-double bond at C-13–C-14 was deduced from the chemical shift of the allylic carbon (C-15, $\delta_{\rm C}$ 17.68). Correlations for H-2′, H₂-5′, and H-7′ to an amide

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⁽⁶⁾ **Perinadine A** (1): colorless amorphous solid; $[\alpha]^{22}_{\rm D} - 33^{\circ}$ (c 1.0, CHCl₃); IR (KBr) $\nu_{\rm max}$ 3405, 3300–2700 (br), 1719, and 1650 cm⁻¹; UV (MeOH) $\lambda_{\rm max}$ 315 (ϵ 1700), 252 (3600), and 215 nm (16 000); CD (MeOH) $\lambda_{\rm ext}$ 313 ($\Delta\epsilon$ -0.5), 257 (+1.3), 224 (-0.6), and 214 nm (+0.6); ¹H and ¹³C NMR, see Table 1; ESIMS m/z 498 (M - H)⁻; HRESIMS (m/z 498.2510 [(M - H)⁻, calcd for C₂₈H₃₆NO₇, 498.2517].

carbonyl carbon (C-6′, $\delta_{\rm C}$ 170.97 and 170.93) suggested that C-7′ was attached to C-2′ and C-5′ through a secondary amide carbonyl group. The existence of a ketone carbonyl ($\delta_{\rm C}$ 206.52 and 206.75) at C-8′ was implied by HMBC correlations for H-7′ and H₂-9′ to C-8′. The relatively lower field chemical shifts for H-2′ ($\delta_{\rm H}$ 5.65 and 5.63) and C-2′ ($\delta_{\rm C}$ 86.75 and 86.87) indicated that another heteroatom in addition to N-1′ was attached to C-2′. The presence of a pyrrolidine ring (N-1′-C-5′) was deduced from the HMBC correlation for H-2′ to C-5′.

The existence of a fully-substituted 2,3*H*-benzopyran skeleton was assigned by HMBC and NOESY correlations as follows. HMBC correlations for H-1 ($\delta_{\rm H}$ 4.24)/C-3 ($\delta_{\rm C}$ 77.68 and 77.58), H-1/C-4a ($\delta_{\rm C}$ 144.57), H-1/C-8a ($\delta_{\rm C}$ 114.26), H-4 ($\delta_{\rm H}$ 2.82)/C-8a, and H₃-10 ($\delta_{\rm H}$ 1.22)/C-4a indicated the presence of 3,4-dimethyl-2,3*H*-pyran ring (C-1-C-4a and C-8a) connected to the pyrolidine ring moiety (N-1'-C-5') at C-1. From the lower field singlet methyl signal at $\delta_{\rm H}$ 2.17 (H₃-11), HMBC correlations to C-4a, C-5 ($\delta_{\rm C}$ 120.11 and 120.15), and C-6 ($\delta_{\rm C}$ 160.98 and 160.99) were observed, thus indicating that C-11 was attached to C-5. The methyl proton signal of C-11 showed NOESY correlations to H₃-10 and a phenol proton ($\delta_{\rm H}$ 12.241 and 12.235), the latter of which was correlated to C-5, C-6, and C-7 ($\delta_{\rm H}$ 99.96) in the HMBC spectrum. This suggested that the phenolic hydroxyl group was connected to C-6. The methine proton of C-1 gave a three-bond correlation for C-8 ($\delta_{\rm C}$ 147.26), which was associated with an ether linkage, and four-bond HMBC correlations to C-5 and C-7. A residual carboxyl carbon (C-12; $\delta_{\rm C}$ 170.78 and 170.75) was suggested to be attached to C-7 because of its relatively higher field chemical shift. The ¹³C chemical shifts of C-6 and C-7 were similar to those of the corresponding carbons in citrinin (4).8,9 The phenolic hydroxyl proton on C-6 appeared as a sharp singlet signal, due to initramolecular hydrogen bonding between the hydroxyl hydrogen atom of C-6 and the carbonyl oxygen atom of C-12. The ether linkage between C-8 and C-2' was suggested by the ROESY correlation for OH-12/H-2' observed at 0 °C. Therefore, the gross structure of perinadine A was assigned as 1.

The relative stereochemistry of the tetracyclic core in perinadine A (1) was elucidated on the basis of NOESY data and $^{1}H^{-1}H$ coupling constants (Figure 2). The $^{1}H^{-1}H$ coupling constant for H-3-H-4 (7.2 Hz) was suggested to be a diaxial orientation for H-3 and H-4. Since a NOESY correlation was observed for H-1/H-3, H-1 was considered to have an axial orientation. The anti relationship between H-1 and H-3' was deduced from NOESY correlations for H-1/H-4' β ($\delta_{\rm H}$ 2.26), H₃-9/H-3', and H-3'/H-4' α ($\delta_{\rm H}$ 2.46), while the NOESY correlation for H-2'/H-3' indicated the cis relationship between H-2' and H-3'. Therefore, the five sp³ methine protons in 1 were concluded to have 1β , 3β , 4α ,

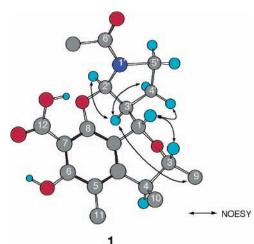


Figure 2. Selected NOESY correlations and relative stereochemistry of the tetracyclic core in perinadine A (1). $^{1}H^{-1}H$ coupling constants (H/H): H-1/H-3′, 4.8 Hz; H-3/H-4, 7.2 Hz; H-2′/H-3′, 5.4 Hz.

 $2'\alpha$, and $3'\alpha$ orientations. The CD spectrum of perinadine A (1) showed exciton maxima at 313 ($\Delta\epsilon$ –0.5), 257 (+1.3), 224 (-0.6), and 214 nm (+0.6), which were similar to the Cotton curve [λ_{ext} 307 ($\Delta\epsilon$ –0.2), 252 (+1.0), 230 (-0.5),

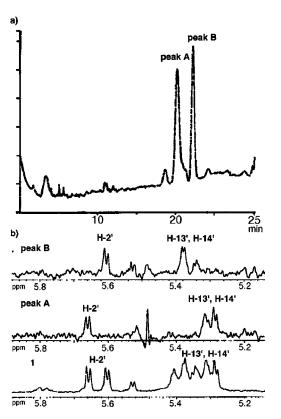


Figure 3. LC NMR analysis of perinadine A (1). (a) HPLC chromatography of perinadine A (1) and (b) part of the ¹H NMR spectra of peaks **A** and **B** and perinadine A (1) in CD₃CN/D₂O/CF₃CO₂H (75:25.0.1).

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⁽⁷⁾ The 13 C chemical shift of C-15 of **1** corresponded to that of C-4 ($\delta_{\rm C}$ 17.6) in *E*-1-chlorobut-2-ene rather than that in *Z*-1-chlorobut-2-ene ($\delta_{\rm C}$ 12.6): Kalinowski, H.-O.; Berger, S.; Braun, S. In *Carbon-13 NMR Spectroscopy*; John Wiley & Sons: Chichester, 1988; pp 293–294.

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and 217 nm (+0.2)] of citrinin (4). This suggested that the three chiral centers in the dihydropyran ring of 1 possessed the same absolute configurations as those of citrinin (4). Therefore, the absolute configurations at C-1, C-3, C-4, C-2', and C-3' were assigned as S, R, R, R, and S, respectively. Two components of perinadine A (1) were examined by LC NMR analysis using C₁₈ HPLC (Figure 3). The HPLC analysis of perinadine A (1) afforded two clearly distinguished peaks **A** (t_R 20.3 min) and **B** (t_R 22.4 min) (Figure 3a). The stopped-flow mode ¹H NMR spectra of peaks A and B disclosed differences in some resonances, especially H-2' signals [peak A, peak B, and perinadine A (1): Figure 3b], suggesting that 1 consisted of two components corresponding to peaks A and B. Nevertheless, these two components would be converted to each other soon after separation and evaporation. In the ¹³C NMR spectrum in MeOH- d_4 , four sets of carbon resonances consisting of two strong and two weak peaks were observed, probably due to the rotational isomer of the N-1'-C-6' amide bond.

Perinadine A (1) is a novel tetracyclic alkaloid with a 2-methyl-3-keto C₁₀ acyl group. Biogenetically, perinadine A (1) may be derived from a known pyrrolidine alkaloid³ (5) isolated from *Penicillium brevicompactum* and citrinin⁸⁻¹⁰ (4), a well-known mycotoxin (Scheme 1). An intermediate a, which may be derived from glutamic acid or proline and a pentaketide, are considered to be converted into 5 by decarboxylation at C-16. Intermolecular cyclization between 5 and an intermediate b equal to citrinin (4) may give perinadine A (1). On the other hand, scalusamide A (2) may be generated from a through reduction of the carboxyl group at C-16, while pyrrolo[2,1-b]oxazines^{3,4} (3) is likely to be converted from 5 through Michael-type cyclization between C-2 and a carbonyl oxygen at C-8.

Perinadine A (1) showed weak cytotoxicity against murine leukemia L1210 cells (IC₅₀, 20 μ g/mL) and antibacterial activity against *Micrococcus luteus* (MIC, 33.3 μ g/mL) and *Bacillus subtilis* (MIC, 66.7 μ g/mL).

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Supporting Information Available: Experimental procedures and spectral data of **1**. This material is available free of charge via the Internet at http://pubs.acs.org.

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⁽¹⁰⁾ Citrinin (4) has not been isolated from the extract of a rotary-shaking cultivation of this strain (N055) of *P. citrinum*. For another strain (N059) of *P. citrinum*, citrinin (4) was observed as a major component in the culture supernatant by stationary cultivation, while this strain produced 4 in rotary-shaking cultivation.